



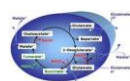
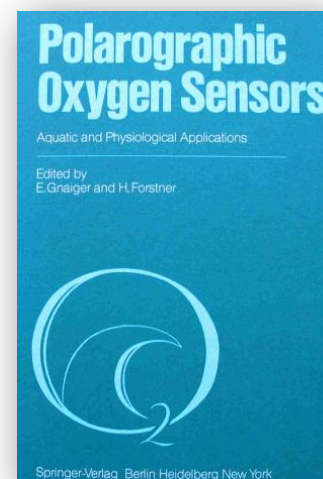
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**O2k-SOP A:** [www.bioblast.at/index.php/MiPNet06.03\\_POS-Calibration-SOP](http://www.bioblast.at/index.php/MiPNet06.03_POS-Calibration-SOP)

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# Polarographic oxygen sensors: calibration, accuracy and quality control SOP

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## A

**Summary:** High-resolution respirometry (HRR; OROBOROS Oxygraph-2k) critically depends on maintenance ([MiPNet19.01B](#)) and accurate calibration of the polarographic oxygen sensors (OroboPOS, POS). Standard operating procedures ([O2k-SOP](#)) are described. (1) Storing and cleaning the O2k-Chambers: clean and ready-to-use O2k-Chambers are a requirement for rapid calibrations and saving time. (2) Quality control for evaluation of proper POS function. (3) Accurate POS calibration ([MiPNet19.01D](#)): A POS responds to partial oxygen pressure,  $p_{O_2}$ . Expressing the signal in terms of  $p_{O_2}$  has several advantages, but respiration is expressed in molar units related to biochemical stoichiometries. Conversion of oxygen partial pressure to oxygen concentration,  $c_{O_2}$ , is based on accurate information on barometric pressure (measured electronically) and oxygen solubilities in experimental media. The oxygen solubility of mitochondrial respiration medium MiR06 relative to pure water (oxygen solubility factor,  $F_M$ ) is 0.92, accurately determined (for MiR05 = MiR06 without catalase; [MiPNet14.13](#)) at 37 °C and 30 °C. At air saturation at standard barometric pressure (100 kPa) and 37 °C, the  $p_{O_2}$  is 19.63 kPa, and the  $c_{O_2}$  is 190.7  $\mu\text{M}$  in MiR06. Absolute errors >10% are common but cannot be accepted in HRR.

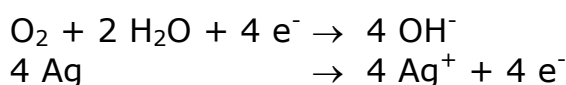
## 1. Polarographic oxygen sensor

Each O2k-chamber is equipped with a polarographic oxygen sensor ([OroboPOS](#), POS). The polarographic oxygen sensor is developed for optimum function of the Oxygraph-2k. The signal is linear in the extended range of partial oxygen pressure of 20 kPa (and up to pure oxygen: 100 kPa) to zero. Thus POS are superior to optical sensors.

The OroboPOS requires minimum service interventions, operates at a high sensitivity and stability for periods of >3 months without change of the membrane at minimum running costs ([MiPNet18.10](#)).

Oxygen diffuses from the sample to the cathode surface through (1) an unstirred layer of the sample at the outer membrane surface, (2) the membrane and (3) the electrolyte layer (Fig. 1). To minimize the unstirred layer of the sample, a high and constant stirring of the sample medium is required. At the cathode the oxygen pressure is effectively held at zero. Under steady-state conditions, the oxygen flux to the cathode depends on the external oxygen pressure, and the electrochemical reduction of oxygen yields an oxygen-dependent consumption of oxygen by the POS which is converted into an electrical signal.

The Clark-type oxygen sensor produces its electrical signal by consuming the oxygen which diffuses across the oxygen-permeable membrane to the cathode (Fig. 1). The cathode and anode reactions are,



At air saturation, the signal of the POS is c. 2  $\mu\text{A}$ . From the stoichiometry (above) and Faraday's law ( $2.591 \text{ pmol O}_2 \cdot \text{s}^{-1} \cdot \mu\text{A}^{-1}$ ), oxygen consumption by the POS at air saturation in a 2  $\text{cm}^3$  chamber is

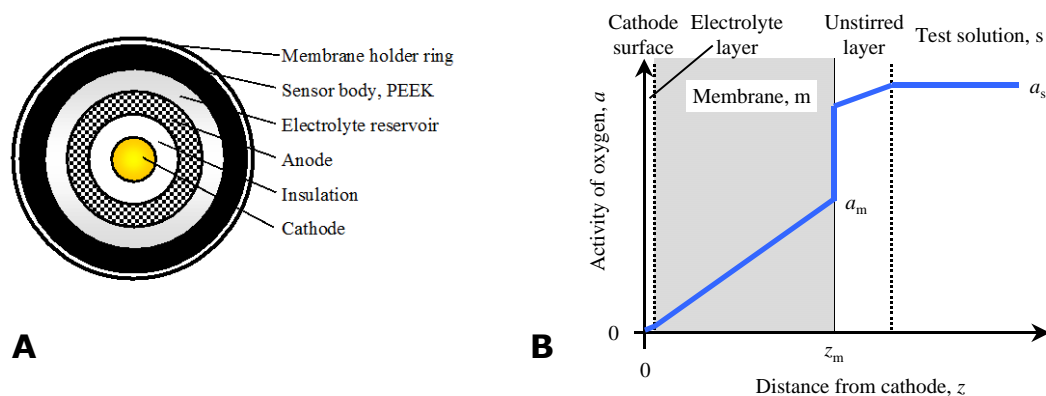
theoretically  $2.6 \text{ pmol}\cdot\text{s}^{-1}\cdot\text{cm}^{-3}$ ), in direct agreement with experiment ([MiPNet14.06](#)).

### 1.1. Cathode

A gold cathode is generally superior to platinum. The sensitivity of polarographic oxygen sensors is a function of cathode area, and long-term stability increases with a high electrolyte volume and a high ratio of anode to cathode area. The signal to noise ratio increases and the relative signal drift at zero oxygen decreases with cathode area. Therefore, the OROBoPOS has a relatively large cathode area (2 mm diameter), yielding a high sensitivity owing to a stable zero current. Signal noise decreases with decreasing oxygen to less than  $\pm 0.003 \text{ kPa}$  (recorded near zero oxygen over 100 data points and 1 s intervals) which is of particular advantage for measurements at physiological intracellular oxygen levels.

### 1.2. Anode

The silver-silver chloride anode has a large area compared to the cathode. The anode may become dark grey-black and is periodically cleaned by treatment with ammonia. Regeneration is possible by a service provided by OROBOROS INSTRUMENTS.



**Figure 1.** The polarographic oxygen sensor (A) consists of a gold cathode and a silver-silver chloride anode, connected by a KCl electrolyte enclosed by an oxygen-permeable membrane. Oxygen diffusion profile (B) at the polarographic oxygen sensor under steady-state conditions in a stirred test solution.

### 1.3. Electrolyte

[KCl solution](#) ( $1 \text{ mol}\cdot\text{dm}^{-3}$ ; 74.56 g potassium chloride per liter distilled water). Dissolve 1.49 g KCl in distilled water to yield a total volume of 20 ml. Before filling the electrolyte into the receptacle of the POS, warm it to c.  $40 \text{ }^\circ\text{C}$  to avoid formation of gas bubbles in the electrolyte reservoir of the POS.

An alkaline electrolyte with KOH did not improve stability of the signal, had no positive effect on the long-term behaviour of the time constant and is less convenient for handling. For these reasons, we do not use a KOH electrolyte.

## A

For a H<sub>2</sub>S insensitive mode of operation at high sulfide concentrations, a special electrolyte should be freshly prepared: Equilibrate distilled water with nitrogen gas. Dissolve 100 g K<sub>2</sub>S·9 H<sub>2</sub>O in 1 liter distilled water. The dissolution requires a long time with automatic stirring. Filter the black precipitate and store in the dark never longer than 6 weeks. The polarizing voltage must be changed from 0.8 V to 0.1 V.

### 1.4. Membrane

At a given oxygen concentration in the test solution, the signal of a POS depends on the properties of the membrane, increasing with diffusion coefficient and oxygen solubility (the product of which is the permeability coefficient), and decreasing with membrane thickness. While a high signal is desirable in terms of a high electronic signal to noise ratio, and a low membrane thickness and high diffusion coefficient increase the time resolution, these advantages are offset by a high background oxygen consumption in the respirometer chamber, an increased sensitivity to the stirring of the sample, and a shortened lifetime of the anode and electrolyte. Therefore, the choice of the membrane requires optimization according to specific requirements. [OroboPOS-membranes](#) (FEP, 25 μm thickness) are used for high-resolution respirometry. Application of a new membrane is simplified by the [OroboPOS-Service Kit](#).



## 2. O2k-Chamber: cleaning and storage with 70% ethanol

### 2.1. Cleaning the chamber (**O2k-SOP**)

Do not exchange stoppers and stirrers between O2k-Chambers, except for [O2k-Troubleshooting](#).

**Clean the chamber before an experiment:** Chemical sterilization is achieved by filling the O2k-Chamber, including the stirrer, and stopper cannula, with 70% ethanol (EtOH). Keep 70% EtOH in the chamber overnight or even for storage over weeks.

1. Filled 70% EtOH up to the top of the chamber holder above the glass chamber, to include the thread in the cleaning.
2. Let the stopper slide to a loosely inserted position.
3. Fill the stopper cups and add the cover slips. EtOH spills over and can be easily siphoned off with the integrated suction system ([ISS](#)).
4. After such storage, washing with distilled water can be done immediately after switching on the Oxygraph-2k.
5. In specific cases (Steininger et al 2002), 3% formaldehyde may be applied for 10 min.

**Clean the chamber after an experiment:** If lipid-soluble inhibitors dissolved in ethanol or DMSO (such as oligomycin,

rotenone, or antimycin A) were titrated, the chamber must be cleaned rigorously with ethanol, since such inhibitor(s) are difficult to be washed out from the chamber and may inhibit mitochondrial respiration in subsequent experiments.

1. Siphon off the cell/tissue/mitochondrial suspension at the end of the experiment.
2. Rinse the chamber with distilled water three times, by filling the chamber up to the rim.
3. Also rinse the surface and cannula of the stopper with distilled water properly.
4. If permeabilized fibres or tissue that may stick to the stirrer is used, it may be necessary to remove the stirrer with a magnetic stick and clean the stirrer mechanically with a clean paper towel. Replace the stirrer into the same O2k-Chamber.
5. Fill the water-cleaned chamber with 70% EtOH.
6. Replace the stopper making sure that the ethanol fills up the receptacle, and leave for 5 min with the clean cover slip covering the stopper.
7. Remove the stopper and siphon off the ethanol to empty the chamber. Repeat these cleaning steps with 70% EtOH three times.
8. Fill the chamber with absolute ethanol (99.6% EtOH).
9. Insert the stopper making sure that the ethanol fills up the cup at the top of the stopper.
10. Replace the cover slip on top of the stopper and leave for 15-20 min.
11. Remove the stopper and place it inverted (with the receptacle at the bottom) on a clean paper towel.
12. Siphon off the ethanol from the chamber and rinse it with distilled water three times. Rinse the stopper by holding it at the receptacle, not at the shaft that fits into the O2k-Chamber to avoid contamination.

## 2.2. Storage in 70% ethanol (**O2k-SOP**)

1. For storage (overnight to several weeks) the O2k-Chamber is filled with 70% EtOH.
2. The stopper is loosely inserted and filled with 70% EtOH up to the rim of the receptacle.
3. A cover slip is placed onto the stopper to minimize evaporation and leakage of EtOH.

» **Goto [Supplement](#)**: EtOH storage saves time and chemicals, and improves wash-out of ethanol-soluble inhibitors from the O2k-Chamber.



## A

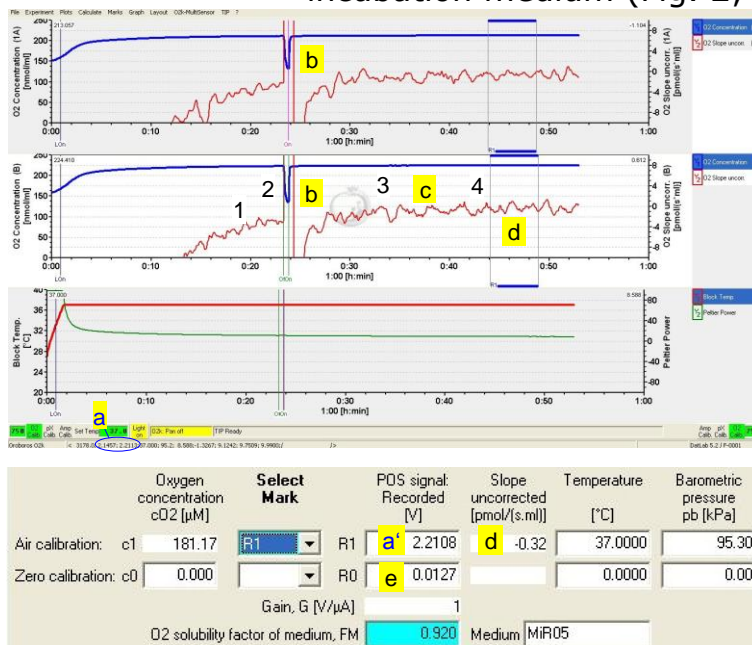
## 3. Calibration and quality control of the OroboPOS (O2k-SOP)

1. Switch on the Oxygraph-2k. Clean the O2k-Chambers. Edit O2k-settings in DatLab and connect.
2. Rise the temperature of experimental medium slightly above experimental before adding at least 2.1 to 2.5 ml medium into each O2k-Chamber, to avoid the formation of gas bubbles and minimize the temperature disturbance.
3. With the stirrer on, insert the stopper fully, check that no air bubbles are contained in the volume-calibrated chamber.
4. Siphon off excess medium from the top of the stopper.
5. Lift the stopper slowly to a position set with the stopper spacer.



**go Bioblast** » [MiPNet19.01D O2k-Calibration](#).

1. About 20 min are required for approximate air equilibration after temperature equilibration of the incubation medium (Fig. 2; time scale is 1 h).



**Quality control a:** The raw signal (blue plot; 1 V = 1 μA at a gain setting of 1) should be close to 1 to 3 V at 25 to 37 °C at sea level up to 1000 m ( $p_b$  101 to 90 kPa). It should be very close to the raw signal obtained at a previous calibration under identical experimental conditions. See O2-Calibration window (a'). At gain setting of 2 the raw signal [V] is multiplied by 2.

**Figure 2.** Quality control of POS performance.

2. Even before final equilibration, perform a stirrer test [F9], switching both stirrers automatically off for 30 s.

**Quality control b:** Upon automatic re-start of the stirrer (On), the increase of the oxygen signal should be rapid and monoexponential (Fig. 3; 30 min time scale).

3. Within 40 min, the oxygen signals should be stable with O2 slope (uncorrected) close to zero.

**Quality control c:** Signal noise should be low, reflected in a noise of the O2 slope (red plot) within  $\pm 2$  ( $\pm 4$  is acceptable)

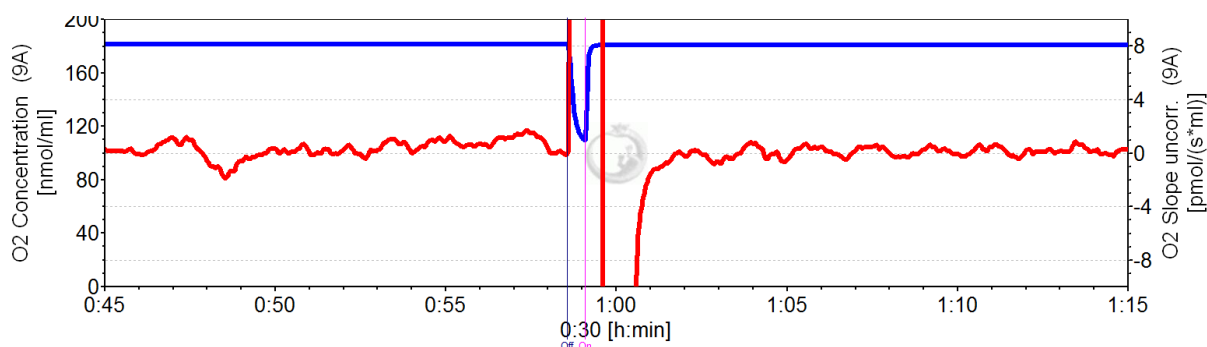
$\text{pmol}\cdot\text{s}^{-1}\cdot\text{ml}^{-1}$  at a data recording interval of 2 s and 40 data points selected for calculation of the slope (Fig. 2).

4. Set a mark on the oxygen signal (R1) and click on O2 Calib. to open the DatLab O<sub>2</sub> calibration window.

**Quality control d:** The slope uncorrected should be within  $\pm 1 \text{ pmol}\cdot\text{s}^{-1}\cdot\text{ml}^{-1}$  averaged across the section of the experiment marked as R1 for air calibration.

5. Continue with an instrumental background test (MiPNet14.06) and perform a zero oxygen calibration.

**Quality control e:** The zero signal at mark R0 for zero calibration should be  $< 2\%$  of R1 (stable at  $< 5\%$  is acceptable).



**Figure 3.** Stirrer test for quality control (standard 30 s) with 30 min time scale displayed with Graph Layout "O2-Background experiment" (MiR05; 37 °C; data recording interval: 2 s; slope smooting: 40 data points). 2014-02-19 P9-01.DLD

## 4. Zero oxygen calibration

### 4.1. Zero calibration with instrumental O<sub>2</sub> background test: TIP2k

**O2k-SOP:** » [MiPNet14.06 InstrumentalBackground](#)

### 4.2. Zero calibration: manual titration of dithionite (**O2k-SOP**)

1. Prepare "zero solution" by dissolving c. 20 mg sodium hydrosulfite (Na-dithionite,  $\text{Na}_2\text{S}_2\text{O}_4$ ; [OroboPOS-Service Kit](#)) or two tips of a spatula in 0.5 ml of water. Dithionite may not work after prolonged storage.
2. With the stirrer on, insert the stopper fully
3. Inject immediately c. 20  $\mu\text{l}$  into the closed O2k-Chamber. A 50  $\mu\text{l}$  Hamilton syringe is recommended.
4. Oxygen depletion is very rapid, zero oxygen is reached within a few minutes, but a few more minutes may be required until a stable "zero" signal is obtained,  $R_0$  [V].
5. Inject again 10  $\mu\text{l}$  zero solution. Repeat as long as the signal responds by a further decline. Siphon off excess medium from the stopper.
6. The zero signal stabilizes quickly ( $< \pm 0.2 \text{ pmol}\cdot\text{s}^{-1}\cdot\text{ml}^{-1}$ ; compare Fig. 3).

A

7. Set a mark over the stable "zero" signal ( $R_0$ ), to complete the two-point oxygen calibration [F5]. Select Mark [R1](#) and Mark [R0](#) for  $R_1$  and  $R_0$ .



**go Bioblast** » [MiPNet19.01D O2k-Calibration](#).

#### 4.3. Zero calibration: mitochondrial respiration

Due to the high oxygen affinity of isolated mitochondria, intact cells and tissue homogenate, residual traces of oxygen are insignificant after respiratory oxygen depletion. Use your experimental sample for such zero oxygen calibration. Alternatively, prepare a stock of bakers yeast, with 200 mg dry yeast in 2 ml physiological salt solution. Stirr heavily to obtain a homogenous suspension of yeast cells and add 50  $\mu$ l yeast suspension into the 2 ml chamber through the cannula of the stopper, using a Hamilton syringe.

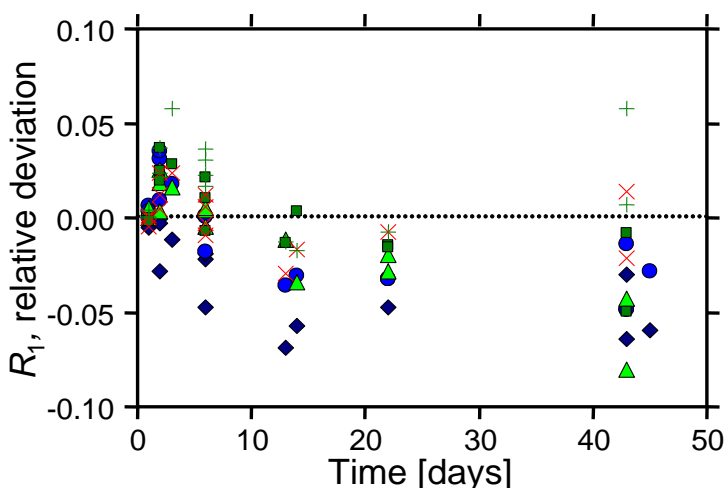


**More details:** [Gnaiger et al \(1995\)](#), [Gnaiger \(2001\)](#).

#### 5. O2-Calibration-List: quality control



**OROBOROS FileFinder:** Click on the icon in the section "O2k-Manual". Go to Chapter 'Oxygen calibration by DatLab' and move to the right to open the Excel file "[O2k-Calibration-List](#)". Save a copy of this Excel Template and paste the calibration parameters into new lines sequentially for chamber (A) and (B), thus generating a data base for quality control of instrumental calibration.



**Figure 4.** Stability of the signals of six OroboPOS at air calibration, R1, over a period of >1 month at constant temperature (25 °C). Membranes were not exchanged and the sensors were left mounted to the O2k-Chambers, which were filled with 70% ethanol during storage, and with mitochondrial respiration medium during calibrations (from Gnaiger 2008).

Trends over time can thus be evaluated (Fig. 4; Gnaiger 2008), and possible irregularities of sensor performance are quickly recognized for intervention ([MiPNet19.01B](#), OroboPOS service).



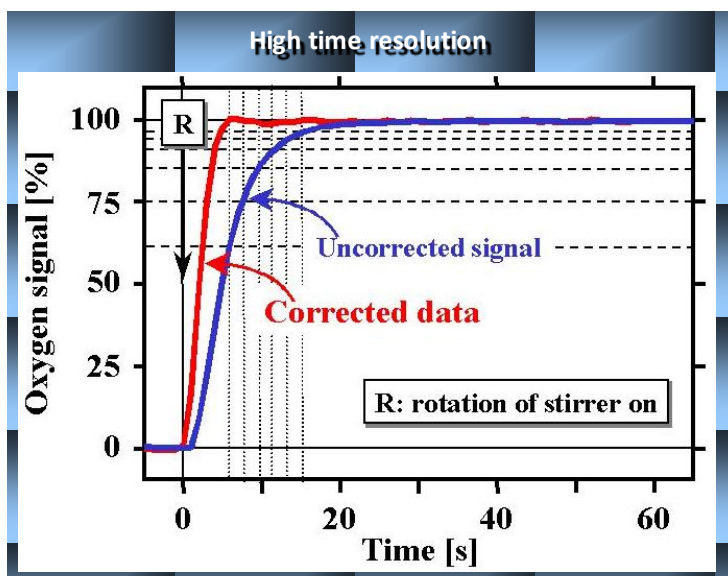
**Full version – go Bioblast**

» [www.bioblast.at/index.php/MiPNet06.03\\_POS-Calibration-SOP](http://www.bioblast.at/index.php/MiPNet06.03_POS-Calibration-SOP)



## Supplement A: Calibration of time constant for signal correction

A



**Figure 5.** Sensors respond with a time delay to rapid changes of oxygen (uncorrected signal). A step change is simply achieved by switching the stirrer off at air saturation, allowing for a local depletion of oxygen at the cathode, followed by switching the stirrer on. The oxygen signal is expressed in % of the total step change. Is the oxygen sensor sufficiently fast for kinetic studies? DatLab yields the answer, gives the exponential time constant (3 s in the present example) and displays the time-corrected data (modified after Gnaiger 2001).

Correction for the time response by using an accurate time constant is essential for high-resolution analysis of kinetic studies, such as ADP pulse titrations and oxygen kinetics involving rapid transitions to anoxia (Gnaiger 2001).

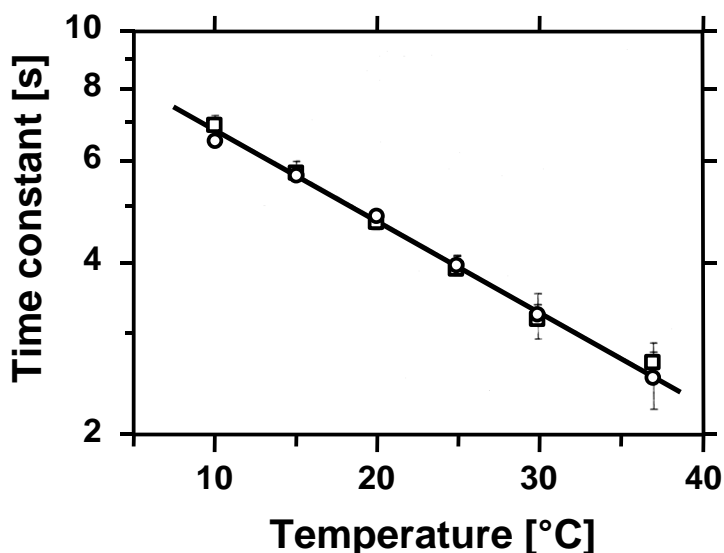
The signal of polarographic oxygen sensors responds with a time delay to rapid changes in the partial pressure of oxygen in the medium (Fig. 5). This convolution of the signal is due to the separation of the oxygen sensor from the experimental medium by a membrane and an electrolyte layer. Consequently, the signal at the cathode responds to a change in oxygen only after

oxygen diffusion has taken place through the membrane to the cathode (Fig. 1B). The time response to changes of  $p_{O_2}$  depends mainly on the thickness of the sensor membrane ( $z_m$ ), the oxygen permeability of the membrane, temperature, and the unstirred boundary layer of the experimental solution (Fig. 1B).

The response time of the oxygen sensor is characterized by an exponential time constant,  $\tau$ . Knowledge of  $\tau$  is crucial both for quality control of the POS and for the time correction of Oxygraph-2k recordings in high-resolution respirometry, particularly in kinetic studies. A fast response of the sensor is indicative of a high quality of sensor maintenance. Prolonged use or storage of the sensor without anode cleaning may increase the response time of the sensor. Such a sensor may be used only if the signal is stable and a high time resolution is not required.

$\tau$  can be experimentally determined by pulse-titration of anoxic into air-saturated medium or by a stirrer test, i.e. turning the stirrer off and on (Fig. 6). Both methods yield identical results. The response is fitted to an exponential function which yields the value of  $\tau$  [s].

A



**Figure 6.** Effect of temperature on the time constant  $\tau$ . The temperature was varied between 10 and 37 °C, and the time constants of both sensors (chamber A and B in the same Oxygraph) were determined by the titration method. Stirring speed 300 rpm; chamber volume 2 cm<sup>3</sup>; titration volume 200-250 mm<sup>3</sup>. Each value represents the mean  $\pm$  SD of 5-6 measurements (from Gnaiger 2001).

$\tau$  depends on experimental temperature, with a  $Q_{10}$  of c. 0.69 (Fig. 6). As expected for a diffusion-controlled process, the time constant  $\tau$  strongly depends on the experimental temperature. A semilogarithmic plot of time constant  $\tau$  vs. temperature results in a straight line (Fig. 6), indicating a 31% decrease in  $\tau$  for a 10 °C increase in temperature.

Stirring speed influences  $\tau$  theoretically only when (1) mixing is slow of the injected (anoxic) solution with the (air-saturated) oxygraph medium (i.e., if the time

constant of the mixing process is in the same range or higher than the time constant of the oxygen sensor), or when (2) unstirred layers (Fig. 1B) play a significant role in oxygen diffusion limitation to the cathode.  $\tau$  is virtually constant between 100 and 700 rpm in anoxic injection experiments, indicating that complete mixing is achieved within a few seconds. A 5% increase of  $\tau$  between 700 and 100 rpm is consistent with the corresponding 5% decrease of the oxygen signal recorded in air-saturated water. This points to more pronounced unstirred layer effects at lower stirring speeds and, at the same time, excludes a significant contribution of the mixing process to  $\tau$ . Similarly, an increase in viscosity associated with the addition of 10% dextran to the experimental medium does not significantly affect the time constant.



**More details:** [Gnaiger \(2001\)](#)

## Supplement B: O2 calibration window in DatLab

A

**Figure 7.** Upon **Show details** [F5] (MiPNet19.01D) oxygen calibration parameters are displayed as calculated by DatLab.

Calibration source	Active file		Calib. POS #	6001		
Oxygen concentration cO2 [ $\mu\text{M}$ ]	<b>Select Mark</b>	POS signal Recorded [V]	Slope uncorrected [pmol/(s.ml)]	Temperature [ $^{\circ}\text{C}$ ]	Barometric pressure pb [kPa]	
Air calibration: c1	180.97	R1	9.7958	0.17	37.0002	95.20
Zero calibration: c0	0.000	R0	0.0278	0.08	37.0007	95.10
Gain, G [V/ $\mu\text{A}$ ]		4				
O2 solubility factor of medium, FM		0.920	Medium	MiR06		
<b>O2 Calibration Info</b>						
<b>Concentration</b>						
Calibration factor for concentration [ $\mu\text{M}/\text{V}$ ]		Fc	18.53	Fc = (c1-c0) / (R1-R0)		
Calibration offset [V]		ac	0.0278	ac = (c1-R0-c0-R1) / (c1-c0)		
<b>Pressure</b>						
Oxygen pressure pO2 [kPa]	POS signal Current I [ $\mu\text{A}$ ]	Oxygen consumption by POS J $^{\circ}\text{O}_2$ (POS) [pmol/(s.ml)]				
Air calibration: p1	18.626	I1	2.4489	I1=R1/G	3.16 J $^{\circ}\text{O}_2$ = 2.591 · (I1-ap) / V	
Zero calibration: p0	0.0000	I0	0.0069	I0=R0/G		
Calibration factor for pressure [kPa/ $\mu\text{A}$ ]		Fp	7.627	Fp = (p1-p0) / (I1-I0)		
Calibration offset [ $\mu\text{A}$ ]		ap	0.0069	ap = (p1-I0-p0-I1) / (p1-p0)		
O2 solubility, SO2 [ $\mu\text{M}/\text{kPa}$ ]	9.72	cO2 = pO2·SO2		O2k Chamber volume, V [ml]		
H2O vapor pressure pH2O* [kPa]	6.27	pO2* = (pb-pH2O*)·0.20946				
Volume fraction of O2 in dry air	0.20946					
Hide details	Copy from file	Cancel		Calibrate and Copy to Clipboard		

→ **Concentration:** Parameters are displayed for conversion of the raw signal to concentration.

**Calibration factor for concentration,  $F_c$  [ $\mu\text{M}/\text{V}$ ]:** This is the multiplication factor,  $F_c$ , calculated to convert the recorded voltage (corrected for the zero signal) into oxygen concentration (Eq. 2).

**Calibration offset,  $a_c$  [V]:** This is the POS zero signal at zero oxygen concentration, which is subtracted from the voltage before multiplication with the calibration factor (Eq. 3).

→ **Pressure:** Parameters are displayed for conversion of the POS signal current to partial oxygen pressure. These are the fundamental parameters for evaluation of signal stability over periods of several months, since the POS responds to partial pressure in the medium rather than concentration.

## A

$p_1$  [kPa],  $p_{O_2}$ : At air saturation,  $p_{O_2}^*$ , a function of temperature and barometric pressure.

$p_0$  [kPa]: Usually zero oxygen concentration, or any other  $p_{O_2}$  at the second calibration point,  $p_0$ .

$I_1 = R_1/G$  [ $\mu A$ ]: POS signal as a current, at air saturation (Eq. 4).

$I_0 = R_0/G$  [ $\mu A$ ]: POS signal as a current, at zero oxygen concentration, or any other other  $p_{O_2}$  at the second calibration point (Eq. 4).

**Oxygen consumption by POS**,  $J^{\circ}_{O_2,POS}$  [ $pmol \cdot s^{-1} \cdot ml^{-1}$ ]: Theoretical oxygen consumption of the oxygen sensor at air saturation under experimental conditions (Eq. 9).

**Calibration factor for oxygen pressure**,  $F_p$  [kPa/ $\mu A$ ]: This is the multiplication factor,  $F_p$ , calculated to convert the current of the POS (corrected for the zero current) into oxygen partial pressure (Eq. 6).

**Calibration offset**,  $a_p$  [ $\mu A$ ]: This is the POS zero current, at zero oxygen pressure, which is subtracted from the current before multiplication with the calibration factor (Eq. 5).

**O<sub>2</sub> solubility** [ $\mu mol O_2 \cdot dm^{-3} \cdot kPa^{-1}$ ]:  $S_{O_2} = c_{O_2} \cdot p_{O_2}^{-1}$ , a function of temperature and oxygen solubility factor of the medium (Eq. 8).

**H<sub>2</sub>O vapor pressure** [kPa]:  $p_{H_2O}^*$ , a function of temperature, is subtracted from the barometric pressure,  $p_b$ .

**Volume fraction of oxygen in dry air**: 0.20946, when multiplied with the pressure ( $p_b - p_{H_2O}^*$ ), it yields the partial oxygen pressure.

**Gain**,  $G$  [V/ $\mu A$ ]: The gain setting (1, 2, 4 or 8 V/ $\mu A$ ) for current to voltage conversion.

**O2k chamber volume**,  $V$  [ml]: The effective aqueous volume of the closed chamber of the Oxygraph-2k.

## Supplement C: Equations for oxygen calibration

### Oxygen concentration and recorded signal

The recorded oxygen signal,  $R_t$ , at experimental time  $t$ , is calibrated in terms of oxygen concentration at time  $t$ ,  $c_{O_2}(t)$ ,

$$c_{O_2}(t) = (R_t - a_c) \cdot F_c \quad (1)$$

where  $F_c$  is the calibration factor based on concentration (Eq. 2),

$$F_c = \frac{c_1 - c_0}{R_1 - R_0} \quad (2)$$

and  $a_c$  is the POS signal at zero oxygen concentration,

$$a_c = \frac{c_1 \cdot R_0 - c_0 \cdot R_1}{c_1 - c_0} \quad (3)$$

$c_1 = c_{O_2}^*$  is the oxygen concentration at equilibrium with air. Typically,  $R_1$  and  $R_0$  are the calibration recordings at air saturation and zero oxygen (if  $c_0 = 0 \mu\text{M}$ , then  $a_c = R_0$ ).

### Oxygen pressure and POS current

In the more general case, the oxygen sensor responds to partial oxygen pressure, and a linear oxygen calibration can be performed at any two calibration pressures of oxygen,  $p_1$  and  $p_0$ . The corresponding oxygen signals in terms of current [ $\mu\text{A}$ ] are  $I_1$  and  $I_0$ . A sensor current of  $1 \mu\text{A}$  yields a raw signal of  $1 \text{ V}$  at a gain setting of  $1 \text{ V}/\mu\text{A}$ .  $G$  is  $2$  or  $4 \text{ V}/\mu\text{A}$  in most O2k applications, and can be changed in the O2k Setup window [F7] to  $1$ ,  $2$ ,  $4$  or  $8 \text{ V}/\mu\text{A}$ . The sensor current,  $I_t$ , at any time  $t$ , therefore, is related to the recorded signal,  $R_t$  [ $\text{V}$ ], according to the gain setting,

$$I_t = R_t/G \quad (4)$$

The zero current or offset,  $a$  [ $\mu\text{A}$ ], is

$$a = \frac{p_1 \cdot I_0 - p_0 \cdot I_1}{p_1 - p_0} \quad (5)$$

If the calibration point  $p_0$  is chosen at zero oxygen concentration, then  $a = I_0$ . The corresponding calibration factor, related to partial pressure and current, is  $F_p$  [ $\text{kPa}/\mu\text{A}$ ],

$$F_p = \frac{p_1 - p_0}{I_1 - I_0} \quad (6)$$

After calibration, comparable to Eq.(1), the partial oxygen pressure,  $p_{O_2}(t)$ , can be calculated from the POS signal current,

$$p_{O_2}(t) = (I_t - a) \cdot F_p \quad (7)$$

### Oxygen concentration and oxygen pressure

The oxygen partial pressure is related to the oxygen concentration,  $c_{O_2}(t)$  [ $\mu\text{M} = \text{nmol}/\text{ml}$ ], by the oxygen solubility,  $S_{O_2}$  [ $\mu\text{M}/\text{kPA}$ ], which is calculated by DatLab on the basis of experimental temperature and the oxygen solubility factor of the experimental medium,  $F_M$ .

$$c_{O_2}(t) = p_{O_2}(t) \cdot S_{O_2} \quad (8)$$



## A

**Oxygen signal and background oxygen consumption**

The oxygen-related POS current,  $I_t - a$  [ $\mu\text{A}$ ] (Eq. 7), results from the steady-state oxygen diffusion from the medium across the membrane and oxygen consumption at the cathode of the POS. Based on the stoichiometry of 4 electrons per molecule  $\text{O}_2$  reduced at the cathode and the Faraday constant (96,485 C/mol), oxygen consumption is expected at  $2.591 \text{ pmol O}_2 \cdot \text{s}^{-1} \cdot \mu\text{A}^{-1}$ . The oxygen consumption by the POS, per volume of the O2k chamber,  $V$  [ml], is  $J^\circ_{\text{O}_2, \text{POS}}$  [ $\text{pmol} \cdot \text{s}^{-1} \cdot \text{ml}^{-1}$ ], calculated as

$$J^\circ_{\text{O}_2, \text{POS}} = 2.591 \cdot (I_t - a_p) / V \quad (9)$$

When the O2k-chamber is closed after equilibration at air saturation, the measured instrumental background oxygen consumption,  $J^\circ_{\text{O}_2}$ , can be compared with this theoretical value. Considering the POS signal at gain 2 and 37 °C to be around 4 V (at gain 4: around 8 V), then  $I_t - a$  is about 2  $\mu\text{A}$  (Eq. 4). At a volume of 2 ml, therefore, the expected instrumental background at air saturation is  $2.6 \text{ pmol O}_2 \cdot \text{s}^{-1} \cdot \text{ml}^{-1}$  (Eq. 9; [MiPNet14.06](#)).

**Supplement D: O<sub>2</sub> solubility and concentration at air saturation****Oxygen pressure and concentration**

It is practical to calculate the saturation concentration for pure water, which then is corrected by the solubility factor of the medium,  $F_M$ , to account for the reduced  $\text{O}_2$  solubility in salt media. Owing to the salting-out effect,  $F_M$  must be  $<1.0$  in salt media used for respiratory studies of mitochondria, cells and tissues.

$F_M$  is typically near 0.9 for Oxygraph media (0.92 for MiR06 and MiR05; [MiPNet14.13](#)). Several oxygen solubilities reported in the literature must be criticized on the basis of physicochemical considerations.

Water in equilibrium with air contains an oxygen concentration proportional to the oxygen solubility and the partial oxygen pressure of air. In the gas-liquid boundary, air is saturated with water vapor at the partial pressure of  $p_{\text{H}_2\text{O}}^*$ . The water vapor pressure is subtracted from the total barometric pressure,  $p_b$ , to obtain the partial pressure of dry air,  $p_b - p_{\text{H}_2\text{O}}^*$ . The volume fraction of dry air is constant at  $\Phi_{\text{O}_2} = 0.20946$ . Therefore, the partial oxygen pressure at air saturation is, for any temperature and barometric pressure,

$$p_{\text{O}_2}^* = (p_b - p_{\text{H}_2\text{O}}^*) \cdot 0.20946 \quad (10)$$

The saturation  $\text{O}_2$  concentration depends on the  $\text{O}_2$  solubility,  $S_{\text{O}_2}$  [ $\mu\text{mol} \cdot \text{dm}^{-3} \cdot \text{kPa}^{-1}$ ],

$$c_{O_2}^* = p_{O_2}^* \cdot S_{O_2} \quad (11)$$

Oxygen solubility is a function of temperature and composition of the medium. In other words, oxygen solubility,  $S_{O_2}$ , is defined as the ratio of partial oxygen pressure and concentration,

$$S_{O_2} = c_{O_2}^*/p_{O_2}^* \quad (12)$$

### Temperature effect on saturation $O_2$ concentration

$p_{H_2O}^*$  (Eq. 10) is the saturation water vapor pressure at experimental temperature.  $p_{H_2O}^*$  is a function of absolute temperature,  $T$  [K], obtained from the experimental temperature,  $\theta$ , recorded in units  $^{\circ}C$ ,

$$T = \theta + 273.15^* \quad (13)$$

The saturation water vapor pressure [kPa] is (Table 1),

$$p_{H_2O}^* = \exp[(-216961 \cdot T^{-1} - 3840.7) \cdot T^{-1} + 16.4754] \quad (14)$$

Until recently, the atm-standard pressure has been used: 1 atm = 760 mmHg = 101.325 kPa. For pure water in equilibrium with air at this atm-standard pressure, the 'unit standard concentration' of oxygen,  $C^*$ , is calculated by the polynomial expression,

$$C^* = \exp\{[(-8.621949 \cdot 10^{11} \cdot T^{-1} + 1.243800 \cdot 10^{10}) \cdot T^{-1} - 6.642308 \cdot 10^7] \cdot T^{-1} + 1.575701 \cdot 10^5\} \cdot T^{-1} - 135.90202 \quad (15)$$

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**Table 1.** Saturation water vapor pressure,  $p_{\text{H}_2\text{O}}^*$ , oxygen pressure,  $p_{\text{O}_2}^*$ , and oxygen concentration,  $c_{\text{O}_2}^*$ , at air saturation and standard barometric pressure,  $p_b^0 = 100$  kPa, in pure water as a function of temperature.  $S_{\text{O}_2}$  is the oxygen solubility, independent of choice of standard pressure.  $f^0$  is the multiplication factor to convert partial  $\text{O}_2$  pressures and concentrations given at atm-standard pressure (1 atm = 101.325 kPa) to the IUPAC standard pressure of 100 kPa (compare Eq. 15),

$$f^0 = (100 - p_{\text{H}_2\text{O}}^*) / (101.325 - p_{\text{H}_2\text{O}}^*)$$

$\theta$ °C	$T$ K	$p_{\text{H}_2\text{O}}^*$ kPa	$p_{\text{O}_2}^*$ kPa	$c_{\text{O}_2}^*$ $\mu\text{mol}\cdot\text{dm}^{-3}$	$f^0$	$S_{\text{O}_2}$ $\mu\text{mol}\cdot\text{dm}^{-3}\cdot\text{kPa}^{-1}$
40	313.15	7.38	19.40	197.6	0.9859	10.18
37	310.15	6.27	19.63	207.3	0.9861	10.56
35	308.15	5.62	19.77	214.2	0.9862	10.83
30	303.15	4.24	20.06	233.0	0.9864	11.62
25	298.15	3.17	20.28	254.8	0.9865	12.56
20	293.15	2.34	20.46	280.4	0.9866	13.70
15	288.15	1.70	20.59	310.9	0.9867	15.10
10	283.15	1.23	20.69	348.1	0.9868	16.83
5	278.15	0.87	20.76	393.9	0.9868	18.97
4	277.15	0.81	20.78	404.3	0.9868	19.46

### Barometric pressure and saturation $\text{O}_2$ concentration

The unit standard concentration and the oxygen concentration at air saturation (Table 1) and actual barometric pressure are related by (compare  $f^0$  in Table 1),

$$\begin{aligned} c_{\text{O}_2}^* &= C^* \cdot p_{\text{O}_2}^* / [(101.325 - p_{\text{H}_2\text{O}}^*) \cdot 0.20946] \cdot F_M \\ &= C^* \cdot (p_b - p_{\text{H}_2\text{O}}^*) / (101.325 - p_{\text{H}_2\text{O}}^*) \cdot F_M \end{aligned} \quad (16)$$

### The barometric altitude relation (BAR)

The partial pressure of oxygen declines with altitude. Hypoxia causes a limitation of maximal aerobic capacity. The  $V_{\text{O}_2\text{max}}$  of acclimatized persons declines at high altitude by c. 11% per 1,000 m, whereas the partial oxygen pressure declines by 12% to 14% per 1,000 m up to 6,000 m, and by 15% to 17% per 1,000 m between 6,000 and 9,000 m. The quadratic model atmosphere equation, MAE, was introduced by John B. West to describe the dependence of average barometric pressure and altitude with high accuracy. An exponential function is the basis of the ICAO Standard Atmosphere, which can be fitted to realistic reference data comparable to the MAE. This leads to the barometric altitude relation, BAR, which

expresses the relationship between barometric pressure,  $p_b$ , and altitude,  $h$  [m], with an even superior fit (Tab. 2):

$$p_b = p_b^\circ \cdot \left(1 - \frac{0.00616 \cdot h}{288.15}\right)^{5.256} \quad (17)$$

The standard pressure at average sea level,  $p_b^\circ$ , is inserted with 101.325 kPa or 760 mmHg to calculate barometric pressure in the respective unit. Compared to the ICAO, only the temperature gradient of  $-6.5$  °C/km (ICAO) was replaced by the parameter  $-0.00616$  °C/m (BAR) which was obtained by a mathematical fit to the reference data in the range of 0 to 9,000 m. 288.15 K is the air temperature of 15 °C at sea level. Deviations between MAE und BAR are less than  $\pm 0.06$  kPa (0.4 mmHg) in the range of 0 to 9 km altitude. In this context the relevance of mitochondrial oxygen kinetics is discussed briefly. The  $p_{50}$  of mitochondrial respiration is 0.01 to 0.1 kPa (0.08 to 0.8 mmHg; this is the partial oxygen pressure at which mitochondrial respiration drops to 50% of maximum values). These generally very low  $p_{50}$  values are important for our understanding of some apparently paradoxical mechanisms of muscular acclimatization and adaptation to hypoxia at extreme altitude (Gnaiger 2013).

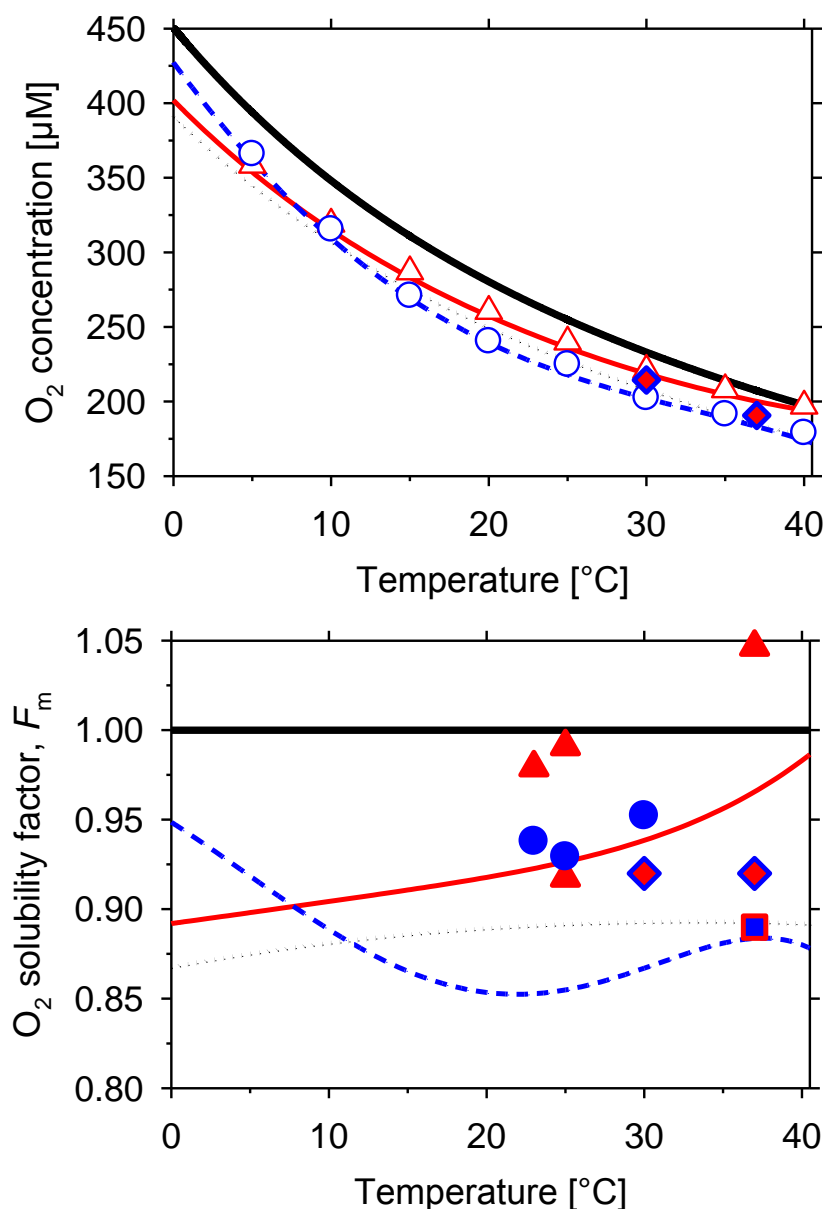
**Table 2.** Barometric pressure,  $p_b$ , and oxygen partial pressure,  $p_{O_2}$ , in dry air and respiratory air saturated by water vapor as a function of altitude,  $h$ . The decline of respiratory air  $p_{O_2}$  is expressed relative to sea level or per 1,000 m change of altitude (from Gnaiger 2013).

$h$ [m]	$p_b$ [kPa]	$p_b$ [mmHg]	Dry air $p_{O_2,da}$ [kPa]	Respiratory air $p_{O_2}$ [kPa]	$p_{O_2}$ [mmHg]	Change rel. to sea level	Rel. change $p_{O_2}/1.000$ m
0	101.3	760	21.2	19.9	149		
1,000	90.4	678	18.9	17.6	132	-0.11	-0.12
2,000	80.5	604	16.9	15.6	117	-0.22	-0.13
3,000	71.5	536	15.0	13.7	103	-0.31	-0.13
4,000	63.3	475	13.3	12.0	90	-0.40	-0.13
5,000	55.9	420	11.7	10.4	78	-0.48	-0.14
6,000	49.2	369	10.3	9.0	68	-0.55	-0.14
7,000	43.2	324	9.1	7.7	58	-0.61	-0.15
8,000	37.8	284	7.9	6.6	50	-0.67	-0.16
9,000	33.0	247	6.9	5.6	42	-0.72	-0.17
575 <sup>a</sup>	94.9	712	19.9	18.6	139	-0.07	
1,675 <sup>b</sup>	83.7	627	17.5	16.2	122	-0.19	
4,559 <sup>c</sup>	59.1	443	12.4	11.1	83	-0.44	
5,240 <sup>d</sup>	54.3	407	11.4	10.1	75	-0.50	
5,364 <sup>e</sup>	53.4	401	11.2	9.9	74	-0.50	
8,848 <sup>f</sup>	33.7	252	7.1	5.7	43	-0.71	

**a:** Innsbruck, A (95.0 kPa; Jul-Aug 2013); **b:** Schröcken, Körbersee, AT (83.6 kPa; Okt 2013); **c:** Monte Rosa, IT (58.4 kPa; Aug-Sep 2004); **d:** Mt Chacaltaya (54.2 kPa; Aug 2012); **e:** Everest Base Camp (52.7 kPa; Mar 2013); **f:** Mt Everest (12, 13). Numbers in parentheses are measurements of  $p_b$  during respirometric studies with the OROBOROS Oxygraph-2k.

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## O<sub>2</sub> solubility factor in salt solutions



**Figure 8.** Oxygen concentration at air saturation and standard barometric pressure (100 kPa; top) and oxygen solubility factor (bottom) in MiR05 (diamonds), KCl medium (open triangles, full line; 150 mmol·dm<sup>-3</sup> KCl) and sucrose medium (open circles, dashed line; 250 mmol·dm<sup>-3</sup> sucrose; data for both media from Reynafarje et al 1985), compared to pure water (upper full line) and 20‰ sea water (lower dotted line). For the parameters of the polynomials see Table 2. The solubility factor for serum is shown by the full square (bottom). Literature data (bottom) on KCl media (closed triangles) and sucrose media (closed circles) show (i) the wide scatter of solubility data, (ii) the erroneous use of values even higher than solubility established for pure water, and (iii) a trend to higher values, particularly in sucrose medium, compared to Reynafarje et al 1985 (see References).

temperature (compare Fig. 8). The solubility factors are compiled in Tab. 5 for different salinities of sea water (Forstner and Gnaiger 1983) and two typical media used with isolated mitochondria (Reynafarje, Costa,

The salting out effect is responsible for the reduced oxygen solubility in aqueous solutions compared to pure water (Fig. 8). Detailed equations are available for calculating the oxygen solubility of sea water at different salinities (Forstner and Gnaiger 1983). Physiological solutions commonly used in Oxygraph studies (Rasmussen, Rasmussen 2003; Reynafarje, Costa, Lehninger 1985) are compared with pure water and 20‰ sea water in Fig. 8. The corresponding polynomial equations are summarized in Tab. 3 for calculating the oxygen saturation concentration in equilibrium with air at various temperatures and standard pressure

(Tab. 4). Characteristic temperatures are commonly used in experimental studies. Under these conditions it is convenient to use oxygen solubility factors for the medium,  $F_M$  (Fig. 8). This factor is independent of barometric pressure, but  $F_M$  changes with



Lehninger 1985). The latter values have been criticized on methodological grounds by Rasmussen and Rasmussen (2003), and the complex temperature dependence of  $F_M$  compared to sea water is doubtful from a thermodynamic perspective Fig. 8).

The oxygen solubility factor of MiR05 (MiR06) is 0.92, at 30 °C and 37 °C (Rasmussen, Rasmussen 2003), corresponding to an oxygen concentration in equilibrium with air under standard conditions ( $c_{O_2}^*$ ) of 214.4 and 190.7  $\mu\text{M}$ , respectively. The oxygen solubility of serum is 9.4  $\text{nmol O}_2 \cdot \text{cm}^{-3} \cdot \text{kPa}^{-1}$  at 37 °C (Baumgärtl and Lübbers 1983). In comparison to the oxygen solubility in pure water (10.56  $\text{nmol O}_2 \cdot \text{cm}^{-3} \cdot \text{kPa}^{-1}$  at 37 °C; Tab. 1), this corresponds to a solubility factor for serum of  $F_M = 0.89$  (Fig. 8) and  $c_{O_2}^*$  of 184.5  $\mu\text{M}$ .

**Table 3.** Parameters of the polynomial fits of oxygen saturation concentration in equilibrium with air at  $p_b^0 = 100 \text{ kPa}$ , for sea water (0‰ and 20‰) and typical Oxygraph incubation media, in the range of  $\theta$  from 5 to 40 °C. Instead of the theoretically based plot of  $\ln(S_{O_2})$  versus  $T^{-1}$ , the fits were performed on the untransformed data, with temperature,  $\theta$ , in units of °C ( $r^2 \geq 0.999$  in all cases). The equation in nested form is,

$$c_{O_2}^* = \{[(b_4 \cdot \theta + b_3) \cdot \theta + b_2] \cdot \theta + b_1\} \cdot \theta + a$$

Medium	A	$B_1$	$b_2$	$b_3$	$b_4$
0‰	450.5946	-12.60381	0.2712233	-0.003808	$2.379 \cdot 10^{-5}$
20‰	390.8769	-10.2165	0.2051415	-0.002746	$1.621 \cdot 10^{-5}$
KCl	401.9152	-10.70002	0.2291496	-0.003283	$2.492 \cdot 10^{-5}$
Sucrose	427.411	-14.4983	0.2762108	-0.0003628	$-3.606 \cdot 10^{-5}$

**Table 4.** Oxygen solubility,  $S_{O_2}$  [ $\mu\text{M} \cdot \text{kPa}^{-1}$ ], for seawater at various salinities (10‰, 20‰, 30‰ and 36‰), and for two typical Oxygraph media (concentrations given in  $\text{mmol} \cdot \text{dm}^{-3}$ ); "Sucrose": 250 sucrose, 5 KCl, 3 K-Hepes, pH 7.05; "KCl": 150 KCl, 3 K-Hepes, pH 7.05.

$\theta$ °C	$S_{O_2}$ for sea water				$S_{O_2}$ for exp. Medium	
	10‰	20‰	30‰	36‰	Sucrose	KCl
40	9.62	9.08	8.58	8.29	8.96	10.01
37	9.98	9.43	8.90	8.61	9.33	10.19
35	10.24	9.67	9.14	8.83	9.54	10.36
30	10.98	10.37	9.80	9.47	10.07	10.90
25	11.86	11.20	10.57	10.21	10.74	11.64
20	12.92	12.19	11.49	11.09	11.70	12.58
15	14.21	13.38	12.59	12.14	13.07	13.75
10	15.79	14.82	13.91	13.39	14.95	15.22
5	17.75	16.60	15.53	14.92	17.42	17.04
4	18.19	17.00	15.89	15.26	17.99	17.45

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**Table 5.** Oxygen solubility factor of the medium,  $F_M$ , for seawater at various salinities (10‰, 20‰, 30‰ and 36‰), and for two typical Oxygraph media (concentrations given in mmol·dm<sup>-3</sup>); "Sucrose": 250 sucrose, 5 KCl, 3 K-Hepes, pH 7.05; "KCl": 150 KCl, 3 K-Hepes, pH 7.05.

$\theta$ °C	$F_M$ for sea water				$F_M$ for exp. Medium	
	10‰	20‰	30‰	36‰	Sucrose	KCl
40	0.945	0.892	0.842	0.814	0.880	0.983
37	0.945	0.893	0.843	0.815	0.884	0.966
35	0.945	0.893	0.844	0.815	0.881	0.956
30	0.945	0.893	0.843	0.815	0.867	0.938
25	0.944	0.892	0.842	0.813	0.855	0.926
20	0.943	0.889	0.838	0.809	0.853	0.918
15	0.941	0.886	0.833	0.804	0.865	0.911
10	0.939	0.881	0.827	0.796	0.889	0.904
5	0.936	0.875	0.819	0.786	0.918	0.898
4	0.935	0.881	0.817	0.784	0.925	0.897

### Supplement E: Storage in 70% ethanol

Experimental tests were performed on storage of the polarographic oxygen sensors (POS) in chambers of the Oxygraph-2k filled with 70% ethanol (EtOH) over periods extended up to several weeks. Based on these results, we recommend to fill the O2k chambers with 70% ethanol for storage over night and over extended periods of time, instead of using distilled water. Storage with EtOH thus replaces the time-consuming procedure described previously, and improves experimental reliability in high-resolution respirometry.

**Until August 2006: 70% ethanol for 20 min:** For many years, we recommended to maintain distilled or deionized water in the O2k chamber during short-term storage over a few days or weeks. This made it necessary to fill the chamber with 70% ethanol for a minimum of 20 min, to obtain a chemically sterilized system.


**Tests and recommendations: extended storage with 70% ethanol:** Intensive tests were carried out which show that the oxygen sensor remains fully functional after storage for several days (weeks) in 70% EtOH. The following considerations led to a new recommendation on using 70% ethanol for short-term storage (20 days) and washing five times with distilled water immediately before addition of mitochondrial respiration medium (experimental salt solution). The test runs have been performed with the PEEK stirrer bars and with new PVDF stirrer bars, and with our titanium stoppers and with the new PVDF stoppers.

1. Time saving: At the end of an experimental day, the chambers are washed with water and then filled completely with 70% EtOH, which remains in the chamber until the next experiment. Then it is not necessary to (1) wait for 20 min upon addition of ethanol, (2) wash the chambers with water, and (3) repeat the 20-min ethanol incubation at the subsequent experimental day. Before the next experiment, the ethanol is simply siphoned from the chamber (ISS), and a chemically sterilized chamber is available.
2. Ethanol saving: Instead of washing with EtOH in the evening and before the next experiment, a single filling of the chamber is sufficient for the O2k chambers and stoppers.
3. Washout of ethanol-soluble inhibitors and uncouplers: Long-term storage with 70% ethanol ensures an extensive solution of trace amounts of inhibitors from the materials of the chamber and stopper into the large volume of EtOH (>5 ml). The stopper is loosely inserted into the chamber, then the receptacle of the stopper is filled up completely and is sealed with the cover slips put on top of the stoppers.

**The following observations provide the basis for the recommendation on ethanol storage:**

1. Over a period of 20 days, the calibration factor of the POS changed by <2% when measured intermittently in salt solution after storage in 70% EtOH.
2. The POS signal stability at air calibration with salt solution corresponded to a slope of  $0.1 \pm 0.3 \text{ pmol}\cdot\text{s}^{-1}\cdot\text{ml}^{-1}$  (mean  $\pm$  SD) in 18 test runs with 6 different sensors over a 20 day period of ethanol storage.
3. Oxygen consumption by the POS at air saturation in salt solution was  $2.0 \pm 0.2 \text{ pmol}\cdot\text{s}^{-1}\cdot\text{ml}^{-1}$  (mean  $\pm$  SD) in 18 test runs (25 °C; 2 ml chamber volume; 6 different chambers) over a 20 day period of EtOH storage.
4. Air equilibration for estimation of the calibration factor was equally rapid after storage in 70% EtOH or in distilled water.
5. The exponential time constant of the POS remained constant over a 20 day period of EtOH storage.
6. The zero current of the POS remained stable over a 20 day period of EtOH storage, when measured in salt solution after oxygen depletion by isolated mitochondria.

## References

- Baumgärtl H, Lübbers DW (1983) Microaxial needle sensor for polarographic measurement of local O<sub>2</sub> pressure in the cellular range of living tissue. Its construction and properties. In: *Polarographic Oxygen Sensors*, Gnaiger E, Forstner H (eds) Springer, Berlin, Heidelberg, New York: 37-65. » 
- Beavis AD, Lehninger AL (1986) Eur J Biochem 158: 315-322.

## A

- Brand MD, Harper ME, Taylor HC (1993) Control of the effective P/O ratio of oxidative phosphorylation in liver mitochondria and hepatocytes. *Biochem J* 291: 739-748.
- Chapell JB (1964) *Biochem J* 90: 225-237.
- Forstner H, Gnaiger E (1983) Calculation of equilibrium oxygen concentration. In: *Polarographic Oxygen Sensors. Aquatic and Physiological Applications*. Gnaiger E, Forstner H (eds) Springer, Berlin, Heidelberg, New York: 321-333. » 📖
- Gnaiger E (2001) Bioenergetics at low oxygen: dependence of respiration and phosphorylation on oxygen and adenosine diphosphate supply. *Respir Physiol* 128: 277-297. » 📖
- Gnaiger E (2013) Die barometrische Höhenrelation (BAR): Form und Funktion. *Jahrbuch 2013 Österr Ges Alpin- Höhenmed*: 51-63.
- Gnaiger E, Steinlechner-Maran R, Méndez G, Eberl T, Margreiter R (1995) Control of mitochondrial and cellular respiration by oxygen. *J Bioenerg Biomembr* 27: 583-596. » 📖
- Hinkle PC, Yu ML (1979) *J Biol Chem* 254: 2450-2455.
- Lemasters JJ (1984) *J Biol Chem* 259: 13123-13130.
- Rasmussen HN, Rasmussen UF (2003) Oxygen solubilities of media used in electrochemical respiration measurements. *Analyt Biochem* 319: 105-113.
- Rettich TR, Battiono R, Wilhelm E (2000) Solubility of gases in liquids. 22. High-precision determination of Henry's law constants of oxygen in liquid water from  $T=274$  K to  $T=328$  K. *J Chem Thermodynam* 32: 1145-1156.
- Reynafarje B, Costa LE, Lehninger AL (1985)  $O_2$  solubility in aqueous media determined by a kinetic method. *Analyt Biochem* 145: 406-418
- Stoner CD (1987) *J Biol Chem* 262: 11445-11453.
- Wilhelm E, Battino R, Wilcock RJ (1977) Low-pressure solubility of gases in liquid water. *Chem Rev* 77: 219-262.

